



## Identify *Lumbricus Rubellus* According DNA Barcoding

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<p><b>Abstract:</b> The species of earthworms is usually determined through careful observation of morphological features, is often accomplished by closely examining morphological characteristics, which are frequently sexual characteristics only seen in mature individuals. Earthworm species identification as such, it is sometimes hard to identify juveniles or cocoons, which might introduce bias into studies that record species richness and abundance. A viable method for species discrimination is DNA barcoding, which uses a brief, standardized DNA fragment for species identification, use of DNA remains the best in determining the species. <i>Lumbricus rubellus</i> have been found on in the common epigeic earthworm from Iraq in Al-Diwaniyah city, the presence of this worm is recorded for the first time in Iraq, specifically in the city of Diwaniyah. This study tested sequence data for the mitochondrial cytochrome c oxidase subunit 1 (COX1) gene in order to identify the utility of DNA barcodes in the identification of earthworm species.</p> <p><b>Keywords:</b> <i>lumbricus rubellus</i>, DNA sequencing, COX1, soil.</p> <p><b>Copyright © 2024 The Author(s):</b> This is an open-access article distributed under the terms of the Creative Commons Attribution <b>4.0 International License (CC BY-NC 4.0)</b> which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited.</p>	<p><b>RESEARCH PAPER</b></p>
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	<p><b>How to cite this paper:</b> Taif Muthher Muslem (2024). Identify <i>Lumbricus Rubellus</i> According DNA Barcoding. <i>Middle East Res J Biological Sci</i>, 4(4): 139-142.</p>
	<p><b>Article History:</b>   Submit: 21.07.2024     Accepted: 20.08.2024     Published: 21.08.2024  </p>

### INTRODUCTION

The first person to refer to earthworms was the scientist Charles Down (1809–1882) before the Geological Society in 1837, and the first to classify earthworms and consider his classification system successful, in the early seventies (Stürzenbaum *et al.*, 2009). Earth worms are the most common creatures in the soil, and belong to the phylum Annelida, classified as clitellata, oligochaeta (Nouri-Aiin *et al.*, 2021). They are particularly widespread in temperate and tropical climates (Kiyasudeen, 2016). From a biomass and activity perspective, earthworms are without a doubt the most significant soil invertebrates in the majority of soil types across the world (Cunha., 2016). Darwin described earthworms as the engineers of the ecosystem (Feller *et al.*, 2003) while Aristotle called them the intestines of the earth (Yadav *et al.*, 2017).

A more precise knowledge of species diversity and its relevance to ecosystem services is needed, as evidence of the sensitivity of soil fauna components to various human activities grows (Norris., 2012) and (Vos *et al.*, 2014). Ignoring cryptic species might cause ecological complexity to be underestimated, which could complicate conservation and biomonitoring efforts (Cahill *et al.*, 2023).

Although *L. rubellus*, the earthworm, is endemic to Western Europe, it is currently considered an

invasive species due to its global proliferation in temperate northern regions (Onrust *et al.*, 2019). *L. rubellus* inhabits soils with a high content of organic matter. Worms like to burrow in soft soil that has enough moisture to allow for exchange gasses, it feeds on decomposing organic matter (Klein *et al.*, 2020).

In most situations, it is hard to identify juveniles of closely related species (such as members of the genus *Lumbricus*) due to their lack of diagnostic characteristics (Richard *et al.*, 2010). Juveniles of closely related species are difficult to identify due to their lack of distinguishing characteristics (Sims *et al.*, 1985). Thus, taxonomists are limited to giving generic identifications for juveniles, which complicates soil investigations aimed at evaluating species richness (Giska *et al.*, 2015).

Among the study's objectives are the following: Molecular diagnosis through DNA extraction, using PCR technology, and relying on DNA sequencing to diagnose earthworms found in the city of Diwaniyah and registering the sample in the Gene Bank.

#### Collecting Samples:

76samples of Earthworm were collected from the soil of three locations in the city of Diwaniyah (Figure 1), located in the north of city 32°00'12.72"N44°52'56.78"E, middle of city 31°59'05.1 "N44°55'21.44' E and south of Diwaniyah city 31°58'5826.9"N44°57'06.72"E, in August 2022.

The samples were collected by digging the soil to a depth of 20-25 cm using a shovel and placed in plastic containers measuring 9\*7\* 16 cm, the forms were sent to the College of Science for the purpose of molecular examination and diagnosis.



Figure 1: Earthworm type *lumbricus rubellus*

## MATERIALS AND METHODS

### DNA Extraction and primer designed

DNA was extracted from earthworms according to the method provided by the company Geneaid Biotech Ltd. USA with a DNA extraction kit gSYNC™. The concentration and purity of the extracted DNA were measured using a Nanodrop.

DNA was extracted by the company Geneaid Biotech Ltd. USA with a DNA extraction kit gSYNC™. The current investigation in order to amplify the mitochondrial COX1 gene (5'TACCGCTCATGCATTCGTAA3') (5'CGCATCCCTCTTCATCGTAT3') producing 357bp fragment, Primers were designed to identify the earthworm *Lumbrica rubellus* by using the NCBI-Genbank (JX531570.1) database and the primer 3 plus online design program.

**PCR:** Upon completion of the polymerase chain reaction method, after that reaction were sent to Bioneer, where the COX1 gene was compared with the COX1 gene sequences found on the NCBI website in order to determine the confirmatory diagnosis of the sample. The results are read according to the Blast program on the NCBI website. Using MEGA6 program, the genetic relationship tree was analyzed. Finally, the sample was registered in the NCBI Genbank submission.

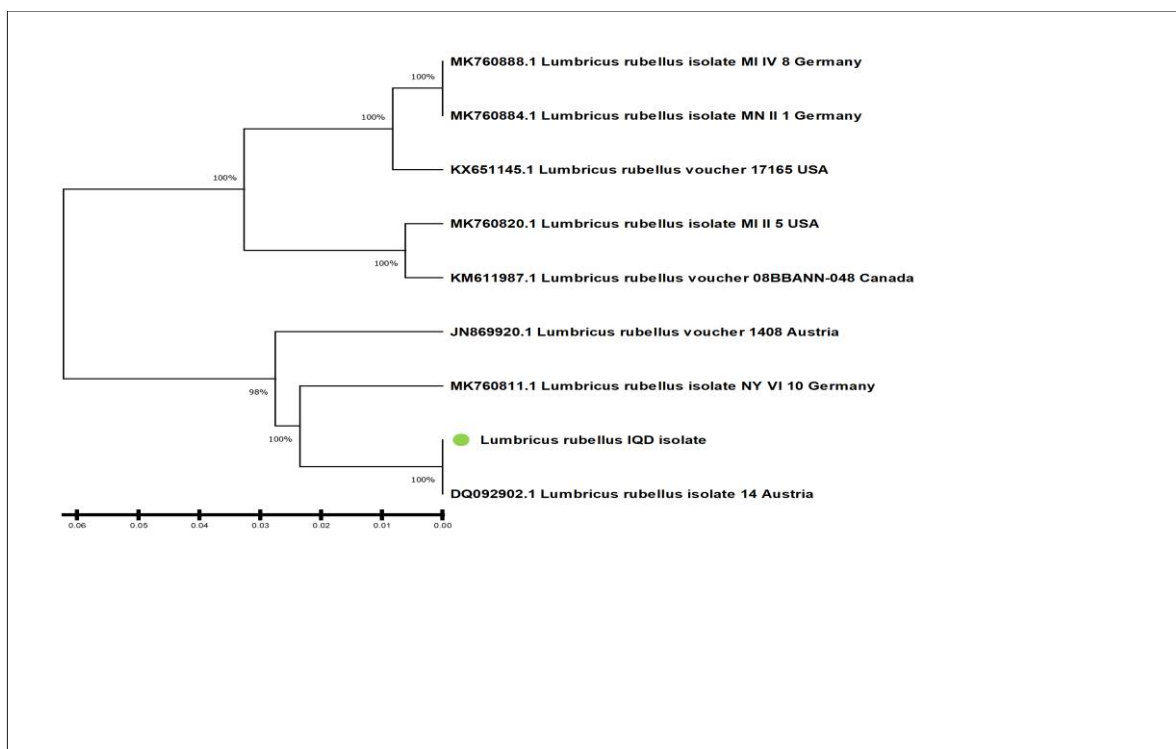
## RESULT AND DISCUSSION

There is difficulty in accurately classifying earthworms depending on the depth of location, size and

color of the earthworm, because some features may be changing depending on the maturity of the earthworm. Therefore, the molecular tool has been used to determine the genus as well as the species of earthworms (Ansari and Saywack.,2011; Jorge Escudero *et al.*, 2022).The results of gel electrophoresis of DNA samples extracted from *Lumbricus rubellus* using the primer for the COX1 gene showed that the DNA bands were (357 bp) and that the marker used was (100-2000pb).The concentration and purity of the extracted DNA was measured using a Nanodrop device, where the DNA concentration was between (177.6 - 829.3) nanograms/microliter, while the purity was between (2.02 - 2.21).

The DNA sequencing method was carried out to genetic identification earth worm species analysis in cytochrome c oxidase subunit I (cox1) gene in local *Lumbricus rubellus* IQD isolate and NCBI-Genbank related country *Lumbricus rubellus* isolates. The phylogenetic tree genetic relationship analysis was showed that The *Lumbricus rubellus* IQD isolate was showed closed related to NCBI-BLAST *Lumbricus rubellus* isolate 14 Austria isolate (DQ092902.1) at total genetic changes (0.06-0.01%) as showed in figure (2). The homology sequence identity between local *Lumbricus rubellus* isolate IQD isolate and NCBI-BLAST *Lumbricus rubellus* isolate isolates were showed genetic homology sequence identity ranged from (88.45 -99.70%). Finally, local *Lumbricus rubellus* isolate were submitted into NCBI Genbank and identified by accession numbers(OP967515).

In a comparative study, a study was conducted in Australia to diagnose earthworms using four mitochondrial genes (12S rRNA, 16S rRNA, COI, and COII) with regard to identifying earthworms, calculating earthworm strains, and discovering cryptic species (Klarica *et al.*, 2012) the study concluded that the four genes are suitable for identifying species, but COI The best gene for coding genes and processing genetic lineages. In a British study, the goal was to examine the genetic structure of earthworm species, *Allolobophora chlorotica*, *Aporrectodea longa*, *Aporrectodea rosea* and *Lumbricus rubellus* all of them comprising highly divergent lineages with species-level divergence in the mitochondrial cytochrome oxidase I (COI) gene Phylogenetic analyses of mitochondrial COI and 16S genes showed the presence of five highly divergent lineages (King *et al.*, 2008). Earthworms have been diagnosed based on one nuclear (H3) and one mitochondrial (COI) marker in northern Europe where *Lumbricus rubellus* and *L. terrestris* and *L. herculeus* have been identified (Martinsson and Erséus., 2017).



**Figure 2: Phylogenetic tree analysis based cytochrome c oxidase subunit I (cox1) gene partial sequence in local *Lumbricus rubellus* IQD isolate that used for genetic species confirmative detection analysis**

Soil contaminations could also affect the distributions of genetic lineages in nature. If the degrees of sensitivity to soil pollutions differ among DNA lineages, some lineage will be lost in polluted spots, which were reducing variation and is consistent with genetic erosion hypothesis. Andre *et al.*, have investigated the highly differentiated population of *L. rubellus* from a Pb-polluted habitat close to Cwmystwyth, UK. The predominant lineages differed by study sites depending on the levels of contamination. This pattern supported the loss of distinct DNA lineage due to pollution. In our research, phylogenetic tree analysis in the least polluted sites. In contrast, it was not found at any of the polluted Al-Diwaniyah city.

In ecotoxicology, earthworm is used for standard toxicity test. The recommended and most commonly used species are *L. rubellus*. However, the taxonomy of these species is not clear because of cryptic diversity. The earthworm *L. rubellus* has been suggested to be a species complex. Römbke *et al.*, reported two distinct DNA clusters of *L. rubellus* that were separated by a distance of 11.2%. Based on the assumption that an uncorrected distance > 10% indicates species level differentiation, these authors hypothesized that *L. rubellus* consisted of cryptic species. This result calls the quality and the comparability of ecotoxicological test into question because cultures of earthworms are rarely barcoded. Nuclear markers were not applied to confirm the DNA clustering of the *L. rubellus* reported by Römbke *et al.*, although previous analysis of nuclear 28S gene indicated possibility that *L. rubellus* from Ireland might be a cryptic species. Therefore, the findings of our

study are particularly relevant because we showed that high DNA divergence, even values exceeding 15%, did not necessarily indicate the presence of cryptic earthworm species. Thus, in addition to crossbreeding experiments, we recommend the use of nuclear data to test for cryptic species in *L. rubellus*.

## CONCLUSION

The study concludes that a viable method for species discrimination is DNA barcoding, which uses a brief, standardized DNA fragment for species identification, use of DNA remains the best in determining the species. *Lumbricus rubellus* have been found on in the common epigeic earthworm from Iraq in Al-Diwaniyah city, the presence of this worm is recorded for the first time in Iraq, specifically in the city of Diwaniyah. This study tested sequence data for the mitochondrial cytochrome c oxidase subunit I (COX1) gene in order to identify the utility of DNA barcodes in the identification of earthworm species.

## REFERENCES

1. Stürzenbaum, S. R., Andre, J., Kille, P., & Morgan, A. J. (2009). Earthworm genomes, genes and proteins: the (re) discovery of Darwin's worms. *Proceedings of the Royal Society B: Biological Sciences*, 276(1658), 789-797.
2. Nouri-Aiin, M., Schall, J. J., Keough, C. A., Wen, Y., & Görres, J. H. (2021). Identifying the unidentifiable: a PCR multiplex protocol for the diagnosis of invasive pheretimoid earthworm

- species, verified by morphological and barcode identification. *Applied Soil Ecology*, 161, 103822.
3. Kiyasudeen S, K., Ibrahim, M. H., Quaik, S., Ahmed Ismail, S., Ibrahim, M. H., Quaik, S., & Ismail, S. A. (2016). General introduction to earthworms, their classifications, and biology. *Prospects of Organic Waste Management and the Significance of Earthworms*, 69-103.
  4. Cunha, L., Brown, G. G., Stanton, D. W., Da Silva, E., Hansel, F. A., Jorge, G., ... & Kille, P. (2016). Soil animals and pedogenesis: the role of earthworms in anthropogenic soils. *Soil Science*, 181(3/4), 110-125.
  5. Feller, C., Brown, G. G., Blanchart, E., Deleporte, P., & Chernyanskii, S. S. (2003). Charles Darwin, earthworms and the natural sciences: various lessons from past to future. *Agriculture, Ecosystems & Environment*, 99(1-3), 29-49.
  6. Yadav, S., & Mullah, M. (2017). A review on molecular markers as tools to study earthworm diversity. *International Journal of Pure and Applied Zoology*, 5(1), 62-69.
  7. Norris, K. (2012). Biodiversity in the context of ecosystem services: the applied need for systems approaches. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 367(1586), 191-199.
  8. Vos, C. C., Grashof-Bokdam, C. J., & Opdam, P. F. M. (2014). *Biodiversity and ecosystem services: does species diversity enhance effectiveness and reliability?* (No. 25). Wettelijke Onderzoekstaken Natuur & Milieu.
  9. Cahill, A. E., Megléc, E., & Chenuil, A. (2024). Scientific history, biogeography, and biological traits predict presence of cryptic or overlooked species. *Biological Reviews*, 99(2), 546-561.
  10. Onrust, J., Wymenga, E., Piersma, T., & Olf, H. (2019). Earthworm activity and availability for meadow birds is restricted in intensively managed grasslands. *Journal of Applied Ecology*, 56(6), 1333-1342.
  11. Klein, A., Eisenhauer, N., & Schaefer, I. (2020). Invasive lumbricid earthworms in North America—Different life histories but common dispersal?. *Journal of biogeography*, 47(3), 674-685.
  12. Richard, B., Decaëns, T., Rougerie, R., James, S. W., Porco, D., & Hebert, P. D. N. (2010). Re-integrating earthworm juveniles into soil biodiversity studies: species identification through DNA barcoding. *Molecular ecology resources*, 10(4), 606-614.
  13. Sims, R. W., & Gerard, B. M. (2023). *Earthworms: keys and notes for the identification and study of the species* (Vol. 31). Brill.
  14. Giska, I., Sechi, P., & Babik, W. (2015). Deeply divergent sympatric mitochondrial lineages of the earthworm *Lumbricus rubellus* are not reproductively isolated. *BMC evolutionary biology*, 15, 1-13.
  15. Ansari, A. A., & Saywack, P. (2011). Identification and classification of earthworm species in Guyana. *International Journal of Zoological Research*, 7(1), 93-99.
  16. Jorge Escudero, G., Lagerlöf, J., Martínez Debat, C., & Pérez, C. A. (2019). Identification of earthworm species in Uruguay based on morphological and molecular methods. *Agrociencia (Uruguay)*, 23(1), 37-46.
  17. Klarica, J., Kloss-Brandstätter, A., Traugott, M., & Juen, A. (2012). Comparing four mitochondrial genes in earthworms—implications for identification, phylogenetics, and discovery of cryptic species. *Soil Biology and Biochemistry*, 45, 23-30.
  18. King, R. A., Tibble, A. L., & Symondson, W. O. (2008). Opening a can of worms: unprecedented sympatric cryptic diversity within British lumbricid earthworms. *Molecular ecology*, 17(21), 4684-4698.
  19. Martinsson, S., & Erséus, C. (2017). Cryptic speciation and limited hybridization within *Lumbricus* earthworms (Clitellata: Lumbricidae). *Molecular Phylogenetics and Evolution*, 106, 18-27.
  20. van Straalen, N. M., & Timmermans, M. J. (2002). Genetic variation in toxicant-stressed populations: an evaluation of the “genetic erosion” hypothesis. *Human and Ecological Risk Assessment*, 8(5), 983-1002.
  21. Römbke, J., Aira, M., Backeljau, T., Breugelmans, K., Domínguez, J., Funke, E., ... & Pfenninger, M. (2016). DNA barcoding of earthworms (*Eisenia fetida*/andrei complex) from 28 ecotoxicological test laboratories. *Applied Soil Ecology*, 104, 3-11.
  22. Andre, J., King, R. A., Stürzenbaum, S. R., Kille, P., Hodson, M. E., & Morgan, A. J. (2010). Molecular genetic differentiation in earthworms inhabiting a heterogeneous Pb-polluted landscape. *Environmental Pollution*, 158(3), 883-890.
  23. Giska, I., Babik, W., van Gestel, C. A., van Straalen, N. M., & Laskowski, R. (2015). Genome-wide genetic diversity of rove beetle populations along a metal pollution gradient. *Ecotoxicology and Environmental Safety*, 119, 98-105.
  24. Pérez-Losada, M., Eiroa, J., Mato, S., & Domínguez, J. (2005). Phylogenetic species delimitation of the earthworms *Eisenia fetida* (Savigny, 1826) and *Eisenia andrei* Bouché, 1972 (Oligochaeta, Lumbricidae) based on mitochondrial and nuclear DNA sequences. *Pedobiologia*, 49(4), 317-324.