

Phage Typing of *Staphylococcus aureus* Isolated from Cancer and Noncancer Patients in Iraq

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Abstract: Background: The bacteriophages have been considerably taken as an important aspect of infectious diseases attacking cancer patients particularly the normal flora causing opportunistic diseases among immunocompromised individuals with tumors. **Materials and Methods:** 400 patients were included and they were 200 patients with cancer diseases and 200 patients complaining of other illnesses. Swabs were taken from nares, sores and wounds and *Staphylococcus aureus* was isolated and identified. The bacteriophage typing was carried out in the Unit of Phages/ Division of Bacteriology/ Central Public Health Laboratory/ Baghdad. **Results:** The present study revealed that 60% (105/175) were typable using the 23 types of bacteriophages which are internationally recognized for lysis of *S. aureus*. The other 40% (70/175) isolates were untypable. It was demonstrated that the most common bacteriophages attacking staphylococci were 42E, 88A, 3C and 71 and the total numbers of *S. aureus* isolates susceptible to these phages were 34, 28, 24 and 22 respectively. The analysis demonstrates marked variability in lysis interaction frequencies among phage types. Phages 42E, 54, and 83A exhibited the strongest lytic interaction frequencies, suggesting high phage compatibility and dominant lysis behavior. Phages 53, 47, and 96 showed weak interaction frequencies, indicating limited lytic association. **Conclusions:** It was found that most of the isolates of *S. aureus* were lysed by the phage group III. A low number of bacterial isolates were lysed by phage group M. It was concluded that 18.7% of the bacterial isolates lysed by phage 29 were susceptible to phage 54 at the same time. It was demonstrated that the most common bacteriophages attacking staphylococci were 42E, 88A, 3C and 71 and the total numbers of *S. aureus* isolates susceptible to these phages were 34, 28, 24 and 22 respectively. The Chi-square test demonstrated a statistically significant difference in the distribution of *Staphylococcus aureus* phage types between cancer and noncancer patients ($p < 0.05$). This indicates that some phage types were more frequently associated with either cancer or noncancer isolates.

Keywords: Cancer Patients, Wounds, *S. Aureus*, Phage Typing.

Research Paper

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INTRODUCTION

The bacteriophages have been considerably taken as an important aspect of infectious diseases attacking cancer patients particularly the normal flora causing opportunistic diseases among immunocompromised individuals with tumors. In a study carried out by Ketchel and Rodriguez [1-4], they concluded that infectious diseases caused almost 20% of deaths among solid tumor patients. Most of the dangerous diseases were caused by Gram-negative bacteria like *Escherichia coli* [5], *Pseudomonas aeruginosa* [6], and *Klebsiella* [7]. Other infectious

diseases of tumor patients were caused by Gram-positive bacteria like *Staphylococcus aureus* and *Staphylococcus epidermis* [7, 8]. These infectious diseases were usually endogenous, community – acquired or hospital-acquired infections [9-11]. The changing of anatomical barriers of cancer patients leads to immunological deficiencies particularly skin and mucous membranes causing invading of the opportunistic normal flora the human body and causing infections for those immunodeficient patients with tumor [12, 13]. Radiation can cause cancer. Highly energetic ionizing radiation (capable of knocking electrons out of atoms and molecules) is the most

hazardous. But non-ionizing radiation such as ultra-violet light, plays an important role in the genesis of human skin cancer. Survivors from Hiroshima and Nagasaki showed a marked elevation in the incidence of leukemia five years after the explosions in 1945. After a later period, the incidence of breast, thyroid, lung, and bowel cancers were found to be modestly elevated [14, 15]. The effect of radiation on man can be observed as weakness, age shortening, redness of the skin, and the elevation of incidence with leukemia [16, 17]. The effect of radiation on human cells appears within 10^{-16} second, usually called the direct physical effect of radiation. The biological effect of radiation not only depends on the dose, but also depends on the relative biological effect (R B E) which is consequently dependent on the type of radiation used. This is defined as the quality factor [16-19]. The effect of the American and affiliated forces attacks using the depleted uranium munitions and the rate of incidence of cancer diseases among the Iraqi peoples by comparing the incidence of cancers before and after the war in 1991[12-21].The present study concerned with the phage typing of most common infectious pathogen disseminated among cancer patients which was *Staphylococcus aureus*. The predominant phage group is belonging to the mixed group. Phage group (II) considered as an epidemiological marker correlated to b-lactamase hyper producer isolates. Staphylococcal isolates particularly MRSA types indicated high prevalence of phage group (II) with highly increase for phage types (Ø3A), related to the skin. Phage types (Ø80/Ø81) played an important roll in Community Acquired Methicillin Resistant *S. aureus* (CAMRSA). Phage groups (M) and (II) were found almost to be integrated for Gentamycin (GN) resistance especially phage type (Ø95) which relatively increased up to 20% in MRSA. Tetracycline (TE) resistant isolates typed by groups (II) and (III). It could be concluded that *S. aureus* isolates from the wound that originated and colonized, and started to build up multi-resistance against the topical treatment [22]. Typing was performed with the 23 phages of the international system: group I, phages 29, 52, 52A, 79, and 80; group II, phages 3A, 3C, 55, and 71; group III, phages 6, 42E, 47, 53, 54, 75, 77, 83A, 84, and 85; and group M (miscellaneous), phages 81, 94, 95, and 96. The methodology described by Blair and Williams was used in accordance with the recommendations of the International Subcommittee for the Phage Typing of *S. aureus* [23, 24]. *S. aureus* strains which were susceptible to phages from two or more of the abovementioned lytic groups were assigned to the mixed-group phage patterns. Strains that could not be typed at 100 times the routine test dilution (100 RTD) were considered non-typeable [25-29]. Most of the studies locally carried out on cancer patients were mostly for survey purposes neglected the epidemiological tools like serotyping, biotyping, resistotyping and phage typing [30, 31].

MATERIALS AND METHODS

Patients

This study was carried out in hospital of atomic medicine, Al-Khansaa Hospital and general hospital of Mosul city, Iraq. 400 patients were included and they were 200 patients with cancer diseases and 200 patients complaining of other illnesses. The acceptance for participation in the present study was taken from all the participants whose native language is Arabic. They were not mentally retarded and they were completely healthy considering hearing and speaking.

Sampling

Swabs were taken from nares, sores and wounds. Swabs were rinsed with sterile nutrient broth as a transport media. 200 swabs of cancer patients and 200 swabs were from noncancer patients [32].

Isolation

All swabs were inoculated on blood and mannitol-salt agars (Oxoid). Inoculated plates were incubated at 37 °C for 24 hours [33].

Identification of *Staphylococcus aureus*

The purified isolates of suspected *S. aureus* were conventionally identified following methods of workers [34, 35].

Bacteriophage Typing

The bacteriophage typing was carried out as described before [21]. This experiment was carried out in the in Unit of Phages/ Division of Bacteriology/ Central Public Health Laboratory/ Baghdad. The set of international phage types used in the present study were: Group I: 29, 52, 52A, 79, 80. Group II; 3A, 3C, 55, 71. Group III: 6, 42E, 47, 53, 54. Group M (miscellaneous): 81, 94, 95, 96.

Statistical Analyses

All statistical analyses were performed using IBM SPSS Statistics for Windows, Version 26.0 (IBM Corp., Armonk, NY, USA). Descriptive statistics such as means, standard deviations, and frequency distributions were computed to summarize the data [36-39].

RESULTS

Phage Typing of *Staphylococcus Aureus*

The present study revealed that 60% (105/175) were typable using the 23 types of bacteriophages which are internationally recognized for lysis of *S. aureus*. The other 40% (70/175) isolates were un-typable. The phage typing used showed that all isolates were human in origin (Table 1). It was found that most of the isolates of *S. aureus* were lysed by the phage group III. A low number of bacterial isolates were lysed by phage group M (Table 2). It was revealed that 61 isolates of *S. aureus* were susceptible to more than one group of phages used. 18.1% of the isolates were lysed by group III and M but

only 7 isolated (6.6%) were lysed by group I and group M (Table 3).

Table 1: Distribution of *Staphylococcus aureus* phage types isolated from cancer and other diseases patients

Source	Phage typed isolates: No.(%)	Non-phage typed isolates: No.(%)	Total isolates
Cancer patients	54 (30.9)	44(25.1)	98(56)
Non-cancer patients	51 (29.1)	26(14.9)	77(44)
Total	105(60)	70(40)	175(100)

Table 2: Distribution of *Staphylococcus aureus* isolates according to their phage typing

Phage group	Phage typed cancer isolates: No.(%)	Phage typed non-cancer patients: No.(%)	Total No.(%)
Group I	12(7.7)	17(11.0)	29(18.7)
Group II	22(14.2)	10(6.5)	32(20.6)
Group III	29(18.8)	38(24.5)	67(43.2)
group M	9(5.8)	18(11.6)	27(18.1)
Total	72(46.5)	83(53.5)	155(100)

Table 3: Distribution of *Staphylococcus aureus* isolates according to their lysis by more than one phage type

Sharing phages	Isolates No. (%)
I+II	9(8.6)
I+III	17(16.2)
I+M	7(6.6)
II+III	9(8.6)
III+M	19(18.1)
Total	61(100)

Frequency of Bacteriophage Types among *Staphylococcus Aureus* Isolates from Cancer and Noncancer Patients

It was demonstrated that the most common bacteriophages attacking staphylococci were 42E, 88A, 3C and 71 and the total numbers of *S. aureus* isolates susceptible to these phages were 34, 28, 24 and 22 respectively (Table 4). Phage type 53 was only able to

lyse one isolate of cancer patient. The Chi-square test demonstrated a statistically significant difference in the distribution of *Staphylococcus aureus* phage types between cancer and noncancer patients ($p < 0.05$). This indicates that some phage types were more frequently associated with either cancer or noncancer isolates (Figure 1).

Table 4: Frequency of *Staphylococcus aureus* phages among isolates from cancer and noncancer patients

Phage type	Cancer patients		Noncancer patients		Total	
	No.	%	No.	%	No.	%
42E	11	20.3	23	45.1	34	32.3
B3A	15	27.7	13	25.5	28	26.6
3C	16	29.6	8	15.6	24	22.8
71	16	29.6	6	11.7	22	20.9
95	5	9.2	14	27.4	19	18.1
29	6	11.1	10	19.6	16	15.2
52	7	12.9	9	17.6	16	15.2
85	7	12.9	8	15.6	15	14.3
54	5	9.2	8	15.6	13	12.3
77	4	7.4	7	13.7	11	10.5
94	3	5.5	7	13.7	10	9.5
81	2	3.7	6	11.7	8	7.6
80	2	3.7	4	7.8	6	5.7
55	4	7.4	2	3.9	6	5.7
3A	5	9.2	1	1.9	6	5.7
B4	3	5.5	1	1.9	4	3.8
96	3	5.5	1	1.9	4	3.8
79	0	0	3	5.8	3	2.8
75	2	3.7	1	1.9	3	2.8
47	1	1.8	2	3.9	3	2.8
53	1	1.8	0	0	1	1.0
Total	118	46.8	134	53.2	252	100

*= Chi-square (χ^2) = 32.06; Degrees of freedom (df) = 20; p-value = 0.0427.

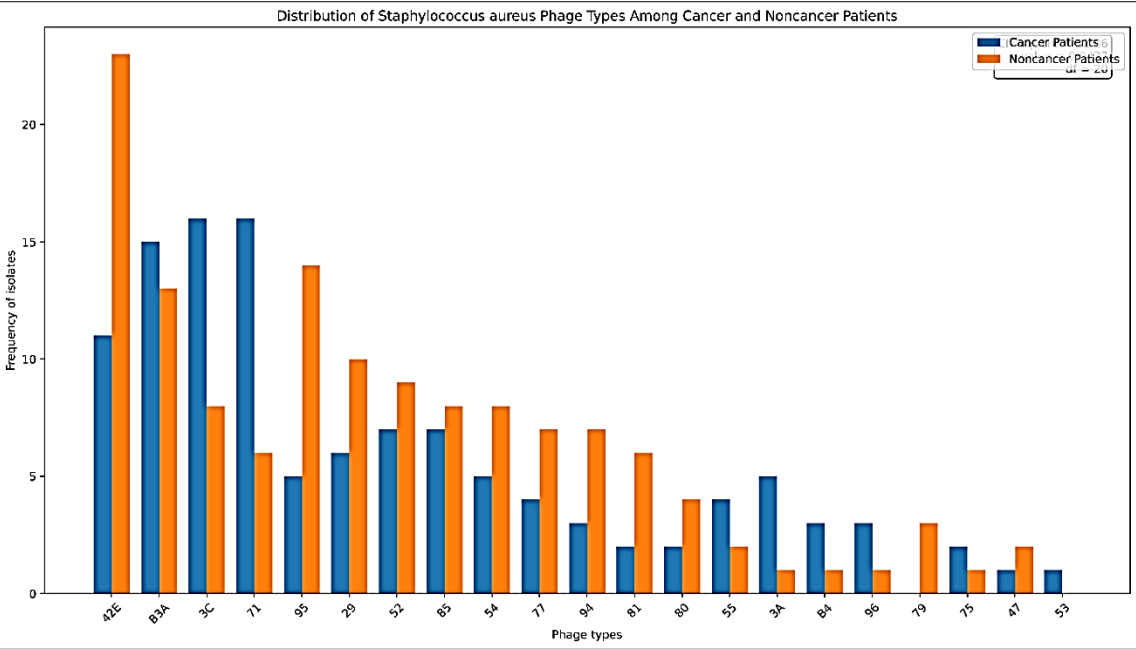


Figure 1: Distribution of *Staphylococcus aureus* Phage Types among Cancer and Noncancer Patients

2x2 Matrix of Phages Causing Lysis to *Staphylococcus Aureus*

The present study revealed that the 2x2 matrix analysis which showed the number of the susceptible isolated of *S. aureus* to 2 phages at the same time (Table 5). The data presented in Table 5 shows that the most common pair of phages was 71/3C that abled to lyse 14 isolates at the same time whereas the two pairs 83A/42E and 83A/77 were able to lyse 9 isolates at the time each. Statistical analyses showed that the mean interaction frequency was 26.35 with standard deviation equal to

16.22. The maximum value was 55 and the minimum was 1. The analysis demonstrates marked variability in lysis interaction frequencies among phage types. Phages 42E, 54, and 83A exhibited the strongest lytic interaction frequencies, suggesting high phage compatibility and dominant lysis behavior. Phages 53, 47, and 96 showed weak interaction frequencies, indicating limited lytic association. The dashed horizontal line represents the overall mean interaction frequency across all phage types (Figure 2).

Table 5: Matrix shows number of *Staphylococcus aureus* lysis at 2x2 pattern of the same time

29	1																		
52	0	7																	
79	0	1	1																
80	0	3	3	1															
3A	0	1	4	0	0														
3C	0	2	6	0	0	5													
55	0	2	4	0	0	2	5												
71	0	1	6	0	0	5	14	5											
54	0	3	3	3	1	3	3	1	2										
53	0	0	0	0	0	0	0	0	0	0									
47	0	0	0	0	0	0	0	0	0	0	1								
42E	0	3	6	3	3	2	3	2	3	7	0	1							
85	1	3	0	1	1	1	2	1	0	4	0	1	3						
84	0	0	0	0	0	0	0	0	0	0	0	0	0	1					
83A	2	3	3	2	1	1	1	1	1	4	0	0	9	8	3				
77	0	2	2	1	1	1	1	0	0	3	0	0	6	4	1	9			
75	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	3	1		
81	1	2	2	2	2	0	0	0	0	3	0	0	6	1	0	6	3	1	
94	2	2	2	2	1	1	0	0	0	3	0	0	5	1	0	3	3	0	5
95	1	3	1	3	2	0	1	1	0	6	0	0	4	2	0	6	2	0	3
	96	29	52	79	80	3A	3C	55	71	54	53	47	42E	85	84	83A	77	75	81
																			94

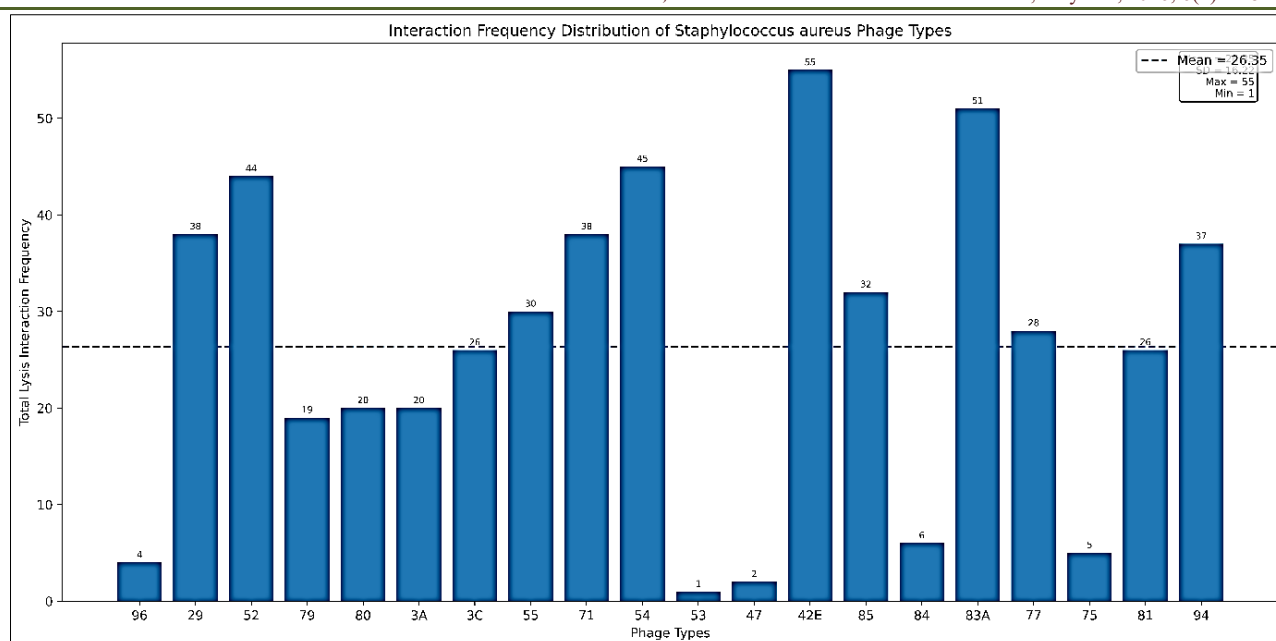


Figure 2: Interaction Frequency Distribution of *Staphylococcus aureus* Phage Types

2x2 Matrix of Percentages of *Staphylococcus Aureus* Cultures Lysed

It was concluded that 18.7% of the bacterial isolates lysed by phage 29 were susceptible to phage 54 at the same time (Table 6). It was noticed also that 23.1% of the isolates that susceptible to phage 54 were lysed by phage 29 at the same Time. It was also concluded that the similarity in frequency of lysis was limited but still existed, e.g. the phage pair 29/52 Frequency was 43.7% in both upper and lower triangles of the matrix. This character is so important for epidemiological Study as a tool for following the distribution of the most common phages that able to lyse the bacterial isolates. The

statistical analyses revealed that mean lysis frequency was 275.71 with standard deviation equal to 107.20. The linear regression coefficient of determinant value was (R^2) value equal to 0.006 with regression P value = 0.7448. The data showed substantial variability in lysis frequencies among different phage types. Phages 54, 83A, 94, and 80 demonstrated the highest lysis activity. The regression analysis indicated a very weak overall linear trend, as reflected by the low R^2 value, suggesting that lysis frequencies vary independently across phage categories rather than following a consistent linear pattern (Figure 3).

Table 6: Frequency matrix of total number of *Staphylococcus aureus* lysis by each phage

	96	29	52	79	80	3A	3C	55	71	54	53	47	42E	85	84	83A	77	75	81	94	95
96	-	25	0	0	0	0	0	0	0	0	0	0	0	25	0	50	0	0	25	50	25
29	6.2	-	43.7	6.2	18.7	12.5	12.5	12.5	2.6	18.8	0	0	18.7	18.7	0	18.7	12.5	0	12.5	12.5	18.7
52	0	43.7	-	6.2	18.7	37.5	25	25	37.5	18.7	0	0	37.5	0	0	18.7	12.5	0	12.5	12.5	6.2
79	0	33.2	33.2	-	33.2	0	0	0	0	100	0	0	100	33.3	0	66.6	33.3	0	66.6	66.6	100
80	0	50	50	16.6	-	0	0	0	0	25	0	0	75	25	0	25	25	0	50	25	50
3A	0	16.6	66.6	0	0	-	83.3	33.3	83.3	50	0	0	33.3	16.6	0	16.6	16.6	0	0	16.6	0
3C	0	8.3	25	0	0	20.8	-	20.8	58.3	12.5	0	0	12.5	8.3	0	4.2	4.2	0	0	0	4.2
55	0	33.3	66.6	0	0	37.3	83.3	-	83.3	83.3	66.6	0	0	33.3	16.6	0	16.6	0	0	0	16.6
71	0	4.5	27.2	0	0	22.7	63.6	22.7	-	9.1	0	0	13.6	0	0	4.5	0	0	0	0	0
54	0	23.1	23.8	23.1	7.7	23.1	23.1	7.7	15.4	-	0	0	53.8	30.8	0	30.7	23.1	0	23.1	23.1	46.2
53	0	0	0	0	0	0	0	0	0	0	-	100	0	0	0	0	0	0	0	0	0
47	0	0	0	0	0	0	0	0	0	0	33.3	-	33.3	33.3	0	0	0	0	0	0	0
42E	0	8.8	17.6	8.8	8.8	5.8	8.8	5.8	8.8	20.6	0	2.9	-	8.8	0	26.4	17.6	0	17.6	14.7	11.7
85	1	20	0	6.6	6.6	6.6	17.3	6.6	0	26.6	0	6.6	20	-	6.6	53.3	26.6	6.6	6.6	13.3	13.3
84	0	0	0	0	0	0	0	0	0	0	0	0	0	25	-	75	25	25	0	0	0
83A	7.1	10.7	10.7	7.1	3.5	3.5	3.5	3.5	3.5	14.3	0	0	32.1	28.5	10.7	-	32.1	10.7	21.4	10.7	21.4
77	0	2	18.2	18.2	9.1	9.1	9.1	9.1	0	0	17.3	0	0	54.5	26.3	9.1	-	9.1	27.2	27.2	18.2
75	0	0	0	0	0	0	0	0	0	0	0	0	0	0	33.3	33.3	3.5	-	12.5	0	0
81	12.5	25	25	25	25	0	0	0	0	37.5	0	0	7.5	12.5	0	75	37.5	12.5	-	50	30
94	20	20	20	20	10	10	0	0	0	30	0	0	50	10	0	30	30	0	50	-	31.5
95	5.3	15.7	5.3	15.7	10.5	0	5.3	5.3	0	30.5	0	0	21.1	10.5	0	31.5	10.5	0	15.8	31.5	-

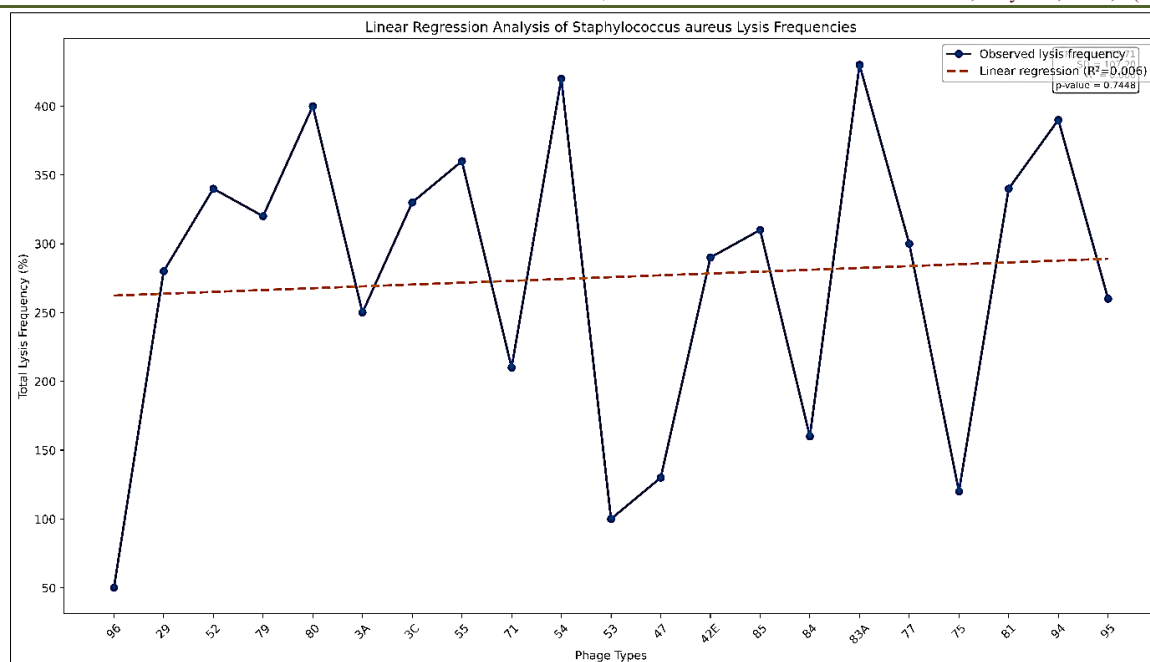


Figure 3: Linear Regression Analysis of *Staphylococcus aureus* Lysis Frequencies

DISCUSSION

The present study revealed that 60% of the *Staphylococcus aureus* isolates were susceptible to phage types used in this investigation [Table 1]. The other 40% isolates were un-typable. At the same time, it was found that phage typing showed that all bacterial isolates were human in origin that is why there was no need to use phage types of animal origin. It was revealed that 61 isolates of *S. aureus* were susceptible to more than one group of phages used. 18.1% of the isolates were lysed by group III and M but only 7 isolated (6.6%) were lysed by group I and group M. The present results were almost similar to those concluded elsewhere [29], who found that 50-80 % of the coagulase – positive staphylococci were able to be typed by one or more of international phages. Khalaf and his co-workers [40], found at Basra district that 63.8% of *S. aureus* isolates were susceptible to the same international phages. On the other hand, Ma *et al.*, [31-41], demonstrated that most of staphylococcal isolates were not susceptible to set of international phage types that used in the present study. MacFarlane and his colleagues [42-44], demonstrated that 19% of the *S. aureus* isolates were resistant to phages and this might be another indication that the source and geographical site of isolates were limitations of bacterial susceptibility to phages. Moreover, it was found that the un-typable isolates of bacteria were under steadily increase in number particularly inside the hospitals, and suggested that this phenomenon might be due to genetical changes of phages to become unable to lyse the bacteria [45-48]. It was found in the present study that most of the isolates were susceptible to group III of the phages used. However, It was found elsewhere that most of the *S. aureus* isolates from Baghdad were highly susceptible phage group I [49], and the results were different compared to the findings concluded in the

present study but almost similar to the patterns demonstrated by MacFarlane and his co-workers [42-51]. It was found also that most of the isolates studied here were susceptible to one or more of the phages belonging to groups I and III. These results were almost similar to those of Khalaf *et al.*, [40]. It was also demonstrated by Williams and Rippon [53], that isolates typeable by more than one group of phages made patterns of epidemiological concern [54-57]. The present study showed that most of the bacterial isolates were susceptible to phages type 42E, 83A, 3C and 71 and these phages were belonging to groups II and III. It was concluded the susceptible isolates to these group are usually associated with wound pus, bacteremia and food-poisoning caused by *S. aureus* [Parker 158]. The frequency of 42E, 83A, 71 and 3C phages lysed isolates of cancer patient. The Chi-square test demonstrated a statistically significant difference in the distribution of *Staphylococcus aureus* phage types between cancer and noncancer patients ($p < 0.05$). This indicates that some phage types were more frequently associated with either cancer or noncancer isolates were mostly different from those of noncancer patients isolates like 42E, 95, 83A and 52. It was found that the phage type 3C was more prevalent in Mosul area compared to other phage types specific for staphylococcal isolates in Basra, Baghdad and Tikrit areas of Iraq [59].

The 2x2 matrix analyses of the phages pair showed that 71/3C pair was the most prevalent causing *S. aureus* isolates lysis at the same time which able to attack 14 isolates. The second most common pairs were 42E/83A and 77/83A which able to lyse 9 isolates by each of them at the same time. Phage type 53 was only able to lyse one isolate of cancer patient. The Chi-square test demonstrated a statistically significant difference in

the distribution of *Staphylococcus aureus* phage types between cancer and noncancer patients ($p < 0.05$). This indicates that some phage types were more frequently associated with either cancer or noncancer isolates. It was tabulated a matrix of percentages of bacterial isolated lysed (Table 6) by dividing the results in Table 5 over data of Table 4. It was noticed that the results of this numerical analyses (Table 6) were not commonly symmetrical, e.g. if you noticed the second line of the matrix you will see that 18.7% of the isolates lysed by phage 29 was lysed by phage 54 at the same time but one can found in line 10 that 23.1% of bacterial isolates which were lysed by phage 54 were lysed by phage 29. Generally, Table 6 shows that the symmetrical pattern was rarely seen, e.g. phage 52/29 showed a symmetrical manner between upper and lower triangles of the matrix when the percentage was 43.7%. This case had a characteristic feature in epidemiological studies for following up these patterns of bacteriophages which are more common with respect to lysis of bacterial isolates. Furthermore, it was found that the most common phage patterns were 52/80, 52A/52, 29/52 and 29/79 with percentages were 78, 87, 68 and 48% respectively [60-63]. The present study showed that the most common phages were 42E, 83A and 3C with frequencies were 32.3, 26.6 and 22.3% respectively. Typing was performed with the 23 phages of the international system: group I, phages 29, 52, 52A, 79, and 80; group II, phages 3A, 3C, 55, and 71; group III, phages 6, 42E, 47, 53, 54, 75, 77, 83A, 84, and 85; and group M (miscellaneous), phages 81, 94, 95, and 96. The methodology described by Blair and Williams [21], was used in accordance with the recommendations of the International Subcommittee for the Phage Typing of *S. aureus* [64, 65]. *S. aureus* strains which were susceptible to phages from two or more of the abovementioned lytic groups were assigned to the "mixed-group" phage patterns. Strains that could not be typed at 100 times the routine test dilution (100 RTD) were considered non-typeable [67]. The resurgence of interest in staphylococcal phages is no longer a peripheral scientific curiosity – it has become a central theme in the broader effort to confront the antimicrobial resistance crisis. From foundational discoveries to current preclinical and clinical developments [68-73], the growing body of evidence supports bacteriophages not only as therapeutic agents but also as powerful tools to understand bacterial evolution, defense systems, and gene transfer. Their versatility extends across medical, veterinary, agricultural, and industrial applications [74-78].

CONCLUSIONS

It was found that most of the isolates of *S. aureus* were lysed by the phage group III. A low number of bacterial isolates were lysed by phage group M. It was concluded that 18.7% of the bacterial isolates lysed by phage 29 were susceptible to phage 54 at the same time. It was demonstrated that the most common bacteriophages attacking staphylococci were 42E, 88A,

3C and 71 and the total numbers of *S. aureus* isolates susceptible to these phages were 34, 28, 24 and 22 respectively. The Chi-square test demonstrated a statistically significant difference in the distribution of *Staphylococcus aureus* phage types between cancer and noncancer patients ($p < 0.05$). This indicates that some phage types were more frequently associated with either cancer or noncancer isolates.

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Statement of Ethics

All the procedures involving human participation were conducted in strict accordance with ethical standards of Institutional Research Committee, Department of Scientific Research, Tikrit University as well as the 1964 Helsinki Declaration and its subsequent amendments or equivalent ethical norms.

Data Availability Statement: The data that support the findings of this study are available from the corresponding author upon reasonable request.

Conflict of Interest Statement: The author declares that he has no conflicts of interest, financial or otherwise.

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REFERENCES

1. Ketchel S, Rodriguez V. Acute infection in cancer patients, seminar. *Oncology*.1978;5(2):167-172.
2. Clokie, M. R. J., Millard, A. D., Letarov, A. V., & Heaphy, S. (2011). Phages in nature. *Bacteriophage*.2011; 1(1): 31–45. <https://doi.org/10.4161/bact.1.1.14942>
3. Rossi CC, Teixeira FCOB, Silva GCD, Pereira MF, Giambiagi-deMarval M. Staphylococcal phages as agents of evolution and innovation: From gene flow to next-generation therapeutics. *Genetic Molecular Biology*. 2026 ; 9:49(suppl 1):e20250131. doi: 10.1590/1678-4685-GMB-2025-0131. PMID: 41677000; PMCID: PMC12895238.
4. Al-Jebouri MM, Al-Meshhadani NS. A note on antibiotic-resistant *Escherichia coli* in adult man, raw sewage and sewage-polluted River Tigris, Nineva. *Journal of Applied Bacteriology*.1985;59:513-518.
5. Bodey GP. Infection in cancer patients.: a continuing association. *The American Journal of Medicine*. 1986; 81:(Supplement 1A):11-26.
6. Griffith SJ. The epidemiology of *Pseudomonas aeruginosa* in oncology patients in a general hospital. *The Journal of Infectious Diseases*.1989;160(6):1030-1036.

7. Klustersky J. Infection in immunocompromised patients I. pathogenesis, etiology and diagnosis. The International Journal of Drug Therapy.1985;8(1):90-99.
8. Al-Jebouri MM(2023) Modellings of infectious diseases and cancers and pollution impacts in Iraq with reference to a novel mathematical model and literature review. Open Journal of Pathology. 13(3):126-139.
9. Al-Jebouri, M M, Sharif, A Y and Abdulla, B A.(1988) The prevalence of antibiotic resistance among three nosocomial pathogens isolated from the maternity hospital in Ninveh. Iraqi Medical Journal, Vol.37,97-101.
10. Eghbalpoor F, Gorji M, Zamani M Moghadam TAM, Genetically engineered phages and engineered phage-derived enzymes to destroy biofilms of antibiotics resistance bacteria, Heliyon.2024;10(15):e35666, <https://doi.org/10.1016/j.heliyon.2024.e35666>.
11. Al-Jebouri MM, Al=Shakarjy. The effect of low-power laser combined with providine-iodine photosensitizer on elastase production of *Pseudomonas aeruginosa* isolated from wounds. Journal of Applied Medical Sciences.2013;2(2):63-67. <http://dx.doi.org/10.4236/oju.2012.23021>
12. Al-Jebouri MM, Al-Ani IA, Al-Jumaily S. The effect of the war of the American and the affiliated forces against Iraq on the distribution and elevation of cancer diseases in Mosul. Consequences of Depleted Uranium used by U.S. and British forces in the 1991 Gulf War Hotel Al-Rashid, Baghdad, Iraq, December 2-3, 1998.
13. Al-Jebouri MM. Clinical Immunology and Allergy. 1st 2dn. Peramerd Publishing Company, Sulaymania, Iraq, 2024,514 p., ISBN:978-9922-9227-7-5, (in Arabic).
14. Currie, G. and Currie, A (1982). Cancer - The Biology of Malignant Diseases. Edward Arnold. London.
15. Al-Jebouri MM(2023) Modellings of infectious diseases and cancers and pollution impacts in Iraq with reference to a novel mathematical model and literature review. Open Journal of Pathology. 13(3):126-139.
16. Raymon M. Nuclear Energy. second edition, Pergamon Press, London,1987.
17. Al-Jebouri MM. A regional study on the infertility of Iraqi males under war impact from 1980 to 2013. World Journal of Pharmaceutical Research. 2015; 4(5): 497-503.
18. Krane K S, Halliday D. Introductory nuclear physics.. Wiley, New York, 1987.
19. Al-Jebouri MM. Medical Bacteriology. 1st edn. Mosul University Press, Mosul,Iraq.1990,388p. (Arabic).
20. Al-Khulaifi MM, Amin M. Nagwa A , Al Salamah AA. Phage typing, PCR amplification for *mecA* gene, and antibiotic resistance patterns as epidemiologic markers in nosocomial outbreaks of methicillin resistant *Staphylococcus aureus*. Saudi Journal of Biological Sciences. 2009; 16:37-49.
21. Blair JE, Williams REO. Phage typing of staphylococci. Bulletin WHO. 1961; 24:771-784.
22. Al-Jebouri MM, Al-Obaidy HS. Providine-iodine as a new lethal photosensitizer for killing of disinfectant-resistant *Staphylococcus aureus* of wounds by light from a helium/neon laser. Proceedings of First Scientific Conference / college of Science 1997; 1,239-250
23. Hood AM. Phage typing of *Staphylococcus aureus*. Journal of Hygiene. 1953;51(1):1-15. doi:10.1017/S0022172400015448
24. Jette LP. Phage Types of *Staphylococcus aureus* Received at the Quebec Public Health Laboratory from 1976 to 1983. Journal of Cclinical Microbiology. 1986;23(1): 180-181.
25. Ma L, Liu Y, Zheng X, Zheng B, Cheng Y, Cai Y, Li Y and Zhang W (2025) Isolation characterization of methicillin-resistant *Staphylococcus aureus* phage SPB against MRSA planktonic cells and biofilm. Front. Microbiol. 16:1554182. doi: 10.3389/fmicb.2025.1554182
26. Chopjitt P, Kanha W, Sachit A, Thongkam J, Kanthain P, Pradabsri P, et al. Bacteriophage-Based Control of Methicillin-Resistant *Staphylococcus aureus*: Anti-Biofilm Activity, Surface-Active Formulation Compatibility, and Genomic Context. Antibiotics (Basel). 2026 Feb 2;15(2):155. doi: 10.3390/antibiotics15020155. PMID: 41750453; PMCID: PMC12937358.
27. Al-Jebouri MM, Mahmood BYR. "Estimation of cytokines involved in acute-phase wound infection with reference to residence time of patients in hospitals". Modern Research of Inflammation. 2019;8(1):1-10. DOI: 10.4236/mri.2019.81001
28. Al-Jebouri MM. The effect of sublethal concentrations of disinfectants on antibiotic resistance pattern of *Pseudomonas aeruginosa*. Journal of Hospital Infection.1989; 14:14-19.
29. Rossi CC, Teixeira FCOB, Silva GCD, Pereira MF, Giambiagi-deMarval M. Staphylococcal phages as agents of evolution and innovation: From gene flow to next-generation therapeutics. Genet Mol Biol. 2026 Feb 9;49(suppl 1):e20250131. doi: 10.1590/1678-4685-GMB-2025-0131. PMID: 41677000; PMCID: PMC12895238.
30. Al-Jebouri MM. "Impact of sublethal disinfectant exposure on antibiotic resistance patterns of *Pseudomonas aeruginosa*". Medical Principles and Practice. 2024;13:1-9. <https://doi:10.1159/000542322>.
31. Ma L, Liu Y, Zheng X, Zheng B, Cheng Y, Cai Y, Li Y and Zhang W (2025) Isolation and characterization of methicillin-resistant *Staphylococcus aureus* phage SPB against MRSA planktonic cells and biofilm. Frontier of Microbiology.2-25; 16:1554182. doi: 10.3389/fmicb.2025.1554182.

32. Al-Jebouri MM, Mahmood BYR. "Estimation of cytokines involved in acute-phase wound infection with reference to residence time of patients in hospitals". Modern Research of Inflammation. 2019;8(1):1-10. DOI: 10.4236/mri.2019.81001
33. Al-Jebouri MM, Mdish SA. Tracing of antibiotic-resistant bacteria isolated from semen of Iraqi males with primary infertility. Open Journal of Urology.2019;9(1):19-29
34. Phillips I. Cowan and Steel's Manual for the Identification of Medical Bacteria. Journal of Clinical Pathology. 1993; 46(10):975. PMID: PMC501641.
35. Al-Jebouri MM, Al-Shakarjy. The effect of low-power laser combined with providine-iodine photosensitizer on elastase production of *Pseudomonas aeruginosa* isolated from wounds. Journal of Applied Medical Sciences.2013;2(2):63-67. <http://dx.doi.org/10.4236/oju.2012.23021>.
36. Al-Jebouri MM, Edham MH. An assessment of biological pollution in certain sector of lower Al-Zab and River Tigris waters using bacterial indicators and related factors in Iraq. Journal of Water Resource and Protection2012, 4:32-38. DOI:10.4236/jwarp.2012.41005
37. Al-Jebouri MM, Trollope DR. The *Escherichia coli* content of *Mytilus edulis* from analysis of whole tissue or digestive tract. Journal of Applied Bacteriology.1991;51(1):135-142.
38. Al-Jebouri MM, Kaki MM. Mathematical Considerations for the Infectious Infertility of Male in Iraq. World Journal of Public Health. 2025; 10(4): 449-458. <https://doi.org/10.11648/j.wjph.20251004.12>
39. Al-Jebouri MM, Kaki MM. Dynamics and Phase Portraits of the SEIQR Model. Annals of Public Health and Epidemiology. 2025; 3(2):1-8.
40. Khalaf FKZ, Al-Hadithi Ht, Mahmood MIA. Phage typing of *Staphylococcus aureus* isolates from food handlers nostrils in Basrah. Iraqi Journal of Microbiology.1989;1(1):7-12.
41. Grdzlishvili N, Lazviashvili D, Kurowska A, Pawlik KJ, Łaczmański , Kakabadze E, et al. Study of the Activity of the Staphylococcus aureus Phage vB_SaS_GE1 Against MRSA Clinical Isolates and Its Impact on the Formation of Dual-Species Biofilms with *P. aeruginosa*. Viruses. 2025;17: 1623.. <https://doi.org/10.3390/v17121623>
42. MackFarlane TW, McGill JC, Samarayke LP. Antibiotic sensitivity and phage typing of *Staphylococcus aureus* isolated from non-hospitalized patients with angular cheilitis. Journal Of Hospital Infection.1984;5:444-446.
43. Al-Jebouri MM, Jasim HH. The relationship between periodontal disease and predisposing factors. Tikrit Journal of Dental Sciences.2016;4(1):68-80.
44. Al-Jebouri MM, Mohamed AA. A study on infertility of males infected with *Mycoplasma hominis* with reference to sperm morphology. Open Journal of Pathology. 2020;11(1):7-21.
45. Naghavi M, Vollset, S.E.; Ikuta, K.S.; Swetschinski, L.R.; Gray, A.P.; Wool, E.E.; Aguilar, G.R.; Mestrovic, T.; Smith, G.; Han, C. Global burden of bacterial antimicrobial resistance 1990–2021: A systematic analysis with forecasts to 2050. Lancet 2024, 404, 1199–1226. [CrossRef]
46. Yung DBY, Sircombe KJ, Pletzer D. Friends or enemies? The complicated relationship between *Pseudomonas aeruginosa* and *Staphylococcus aureus*. Molecular Microbiology. 2021; 116: 1–15. [CrossRef]
47. Al-Jebouri M.M, Kaki MNM. "Application of Matrices Modelling for Infectious Diseases of Humans" Open Journal of Applied Sciences; 2025;15: 2733-2758. <https://doi.org/10.4236/ojapps.2025.159184>
48. Sultan HI, Al-Jebouri MM. Pulmonary tuberculosis in Al-Zab district. Tikrit Medical Journal. 2010;16(1):37-41
49. Al-Rawi F, Al-Salihi SMSF, Abdulla T, Al-Awad T, Sarmin MA. Coagulase positive Staphylococci phage typing in Iraq. Scientific Journal of Nursing.1985; 1:27.
50. Wang L, Tkhilaishvili T, Trampuz A, Gonzalez Moreno M. Evaluation of staphylococcal bacteriophage Sb-1 as an adjunctive agent to antibiotics against rifampin-resistant *Staphylococcus aureus* biofilms. Frontiers of Microbiology. 2020;11: 602057.. [CrossRef]
51. Verheul M, Mulder A A, van Dun S C, Merabishvili M, Nelissen R G, de Boer M G, et al. Bacteriophage ISP eliminates *Staphylococcus aureus* in planktonic phase, but not in the various stages of the biofilm cycle. Science. Report. 2024;14: 14374.
52. Al-Jebouri MM, Al-Janabi M, Ismail H. 2013. The prevalence of toxoplasmosis among female patients in Al-Hawija and Al-Baiji districts in Iraq. Open Journal of Epidemiology. 2013;3(1):1-4.
53. Williams R Eo, Rippon JE. Bacteriophage typing of strain of *Staphylococcus aureus* from various sources. Lancet. 1953;1:510-514.
54. Glonti T.; Pirnay J P. In Vitro Techniques and Measurements of Phage Characteristics That Are Important for Phage Therapy Success. Viruses. 2022; 14: 1490.. [CrossRef]
55. Jakočič, D, Moodley A A .rapid bacteriophage DNA extraction method. Methods Protocol. 2018; 1: 27. [CrossRef]
56. Al-Jebouri MM, Al-Dobony HM. Incidence of antibiotic-resistant bacteria in urine from healthy secondary school girls in Mosul. Iraqi Medical Journal.1985;33:97-107.
57. Al-Jebouri MM, Madish SA. The role of sex hormones in the formation of renal stones with reference to urinary tract infections of Iraqi patients. World Journal of Pharmacy and Pharmaceutical Sciences. 2014;3(4):352-365.

58. Parker MT. The significance of phage-typing patterns in *Staphylococcus aureus* in: Easman,C.S,F, and Adlam C.edt. Staphylococci and staphylococci infections. Academic Press WC. New York. Pp.33-60.
59. Rahman A,GY. The study of enterotoxins produced by *Staphylococcus aureus* from different groups and clinical material in Mosul. M.Sc. Thesis, University of Mosul (in Arabic).
60. Trizna E Y, Yarullina M N, Baidamshina D R, Mironova A V Akhatova F S , Rozhina E V et al. Bidirectional alterations in antibiotics susceptibility in *Staphylococcus aureus*—*Pseudomonas aeruginosa* dual-species biofilm. Sci. Rep. 2020, 10, 14849. [CrossRef]
61. Kebriaei R, Lehman S M, Shah R M, Stampe K C, Coyne A J K, Holger D, et al. Optimization of Phage-Antibiotic Combinations against *Staphylococcus aureus* Biofilms. Microbiol. Spectr. 2023, 11, e04918-22. [CrossRef]
62. Al-Jebouri MM, Mdish SA. Tracing of Sortoli-cell-only syndrome and other histopathological abnormalities in Iraqi males with primary infertility of azoospermia. Open Journal of Pathology. 2019;9(1):10-17.
63. Al-Jebouri MM, Al-Rahaley IM. An assessment of antibiotic resistance and resistotyping of *Escherichia coli* from stools of infants. Journal of Chemotherapy. 1991;3:119-121.
64. Al-Jebouri MM, Al-Alwani HR. Angiotensin I-converting enzyme gene polymorphism in patients with chronic renal failure. World Journal of Pharmaceutical Research. 2014;1-11.
65. Al-Jebouri MM, Mahmood BY. Prevalence of different types of microorganisms and levels of complement C5a in patients with acute-phase wound infections. Open Journal of Pathology.2019;02-19.
66. Al-Jebouri MM, Hasen AH. Vitamin D3 variation between children and adults with reference to renal stone, environment and urinary tract infections. Open Journal of Urology. 2012; 2(3):119-126.
67. Barber, K.E.; Shammout, Z.; Smith, J.R.; Kebriaei, R.; Morrisette, T.; Rybak, M.J. Biofilm time-kill curves to assess the bactericidal activity of daptomycin combinations against biofilm-producing vancomycin-resistant *Enterococcus faecium* and faecalis. Antibiotics2021, 10, 897. [CrossRef]
68. Becker K, Both A, Weißelberg S, Heilmann C and Rohde H . Emergence of coagulase-negative staphylococci. Expert Review of Antimicrobial Infections Therapy.2020; 18:349-366.
69. Bernheim A, Sorek R. The pan-immune system of bacteria: Antiviral defence as a community resource. Natural Review of Microbiology.2020;18:113-119.
70. Al-Jebouri M M, Al-Faham Y M N. Synergistic effect of laser irradiation and photosensitizers on *Pseudomonas aeruginosa* isolated from the wound. World Journal of Pharmacy and Pharmaceutical Sciences.2025;14(11): 502-517.
71. Al-Jebouri MM, Al-Jebouri OAH. Prevalence, Incidence, and Associated Risk Factors of Urolithiasis among Population of Salah-Aldeen Province In Iraq. World Journal of Pharmaceutical Research.2026; 15(5), 817–832.
72. Al-Jebouri MM, Al-Jebouri OAH. Urinary Metabolic Profile and Stone Composition, Location and Recurrence in Kidney Stone Formers of the Salah-Aldeen Province, Iraq. Middle East Research Journal of Microbiology and Biotechnology., 2026 ; 6(1): 1-9.
73. Al-Jebouri MM, Al-Jebouri OAH. Antibiotic Resistance Patterns of Bacteria Isolated from Patients with Urolithiasis in Tikrit City, Iraq. World Journal of Pharmacy and Pharmaceutical Sciences.2026; 15(5): 296-312.
74. Carroll KC, Burnham CAD and Westblade LF .Clinical importance of *Staphylococcus* From canines to humans: pseudintermedius. PLoS Pathology.2021; 17:e1009961.
75. Carvalho A, Giambiagi-deMarval M, Rossi CC . Mammaliococcus sciuri's pan-immune system and the dynamics of horizontal gene transfer among Staphylococcaceae: A One-Health CRISPR tale. Journal of Microbiology.2024; 62:775-784.
76. Al-Jebouri MM, Anton BG. *Escherichia Coli* Type-I Isolated From Mid-Fingers As Indicator A New Of Potential Microbial Health Hazards Associated With Food Handler Employees. World Journal of Pharmacy and Pharmaceutical Sciences.2026; 15(2), 1147–1164.
77. Al-Jebouri MM, Kaki MM, Mutlek T. Quantitative assessment of wound healing of mice treated with combinations of laser irradiations and antibiotics via experimental mathematical modelling. 2026; 15(2): 1165-1177.
<https://doi.org/10.11648/j.wjph.20251004.12>
78. Al-Jebouri MM, Al-Bayati HS. Effect of Disinfectants Exposure on the Susceptibility of Antibiotics, Disinfectants, Heavy Metals and Biochemical Profile for *Staphylococcus aureus* Isolated from Caesarean Wounds” Middle East Research Journal of Microbiology and Biotechnology. 2026 ; 6(1): 22-34.