

Middle East Research Journal of Medical Sciences ISSN: 2789-7699 (Print) & ISSN: 2958-2024 (Online) Frequency: Bi-Monthly DOI: 10.36348/merjms.2024.v04i01.006



How Are Short-Chain Fatty Acids Associated with Deranged Lipid Profiles in Cad Patients?

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Abstract: Background: The phenomenon of Coronary Slow Flow (CSF) is an	Research Paper
angiographic clinical verity, which is characterized by late opacification of the distal	*Corresponding Author:
segments of the coronary artery without having significant stenosis. However, a definite	Dr. Anil Batta
and reliable mechanism of CSF is still not known. Short-chain fatty acids (SCFA) such	Professor & Head, Department of
as acetate, butyrate, and propionate are produced as a result of the fermentation of	Biochemistry, Muzaffarnagar Medical College, Muzaffarnagar
indigestible dietary fibres in the gut by the microbiota. Many studies have investigated	How to cite this paper:
the role of SCFA as a related signalling pathway in inflammation, glucose metabolism,	Anil Batta (2024). How Are
and lipid metabolism. In this study, we investigated the correlation between Short Chain	Short-Chain Fatty Acids
Fatty Acids and Lipid Profile serum in Patients with Slow Flow Coroner. <i>Materials and</i>	Associated with Deranged
Methods: 50 patients who were referred to the Department of Internal Medicine,	Lipid Profiles in Cad
Muzaffarnagar Medical College, Muzaffarnagar, were selected based on inclusion and	Patients?. Middle East Res J.
exclusion criteria. Data was obtained through laboratory examination and stool samples.	Med. Sci, 4(1): 31-35
Stool samples were analyzed for SCFA (acetate, propionate, and butyrate acids) with gas	Article History:
chromatography. <i>Results</i> : The results of the present study indicate that SCFA, acetate,	Submit: 19.12.2023
propionate, and valerate did not show a significant correlation with lipid profile (P>0.05).	Accepted: 24.01.2024
The level of fecal butyrate was negatively correlated with HDL ($p=<0.05$; $r = -0.532$).	Published: 09/02/2024
Conclusions: Our study indicated that the level of butyrate was a moderate negative	
correlated with HDL inpatient with slow flow coroner.	

Keywords: SCFA, Lipid Profile, coronary slow flow. **Copyright © 2024 The Author(s):** This is an open-access article distributed under the terms of the Creative Commons Attribution **4.0 International**

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INTRODUCTION

The phenomenon of Coronary Slow Flow (CSF) is an angiographic clinical verity, which is characterized by late opacification of the distal segments of the coronary artery without having a significant stenosis [1]. Small vessels, diffuse atherosclerosis, vascular inflammation, endothelial dysfunction, and platelet aggregation dysfunctions are some of the theories that have been proposed to explain the pathophysiology of CSF [2, 3]. The gut microbiota has been demonstrated to have a key role in the progression of atherosclerosis, but the exact mechanism remains unknown. Short-chain fatty acids (SCFAs), such as acetate, propionate, and butvrate, are metabolites created by bacterial fermentation in the colon from otherwise indigestible polysaccharides (fibres) [4]. In rodents and humans, SCFAs have been found to reduce plasma concentrations of cholesterol. Propionate is being considered as a possible contender for decreasing plasma cholesterol levels; however, the outcomes of trial evaluation are controversial [5, 6].

MATERIALS AND METHODS

Patients and Study Design

This cross-sectional study included twelve patients with continuing episodes of chest pain and angiographic criteria for the CSF were enrolled. The present study was conducted from July 2021 to December 2021 in the Department of Biochemistry, Muzaffarnagar Medical College, Muzaffarnagar, India Cardiac Catheterization Laboratory. and A11 angiographic examinations were conducted by two cardiologists who were blinded to the clinical characteristics of the patients and assessed the flow in coronary arteries using the Thrombolysis in the Myocardial Infarction (TIMI) frame count method, described by Gibson et al., [7]. Individuals who had active antibiotic treatment or within the month prior to angiography, yogurt consumption or laxative medicine for the last four weeks or who had undergone surgery for intestinal tumours were excluded from the study. The whole blood was analyzed for fasting blood sugar (FBS), total cholesterol (TC), triglycerides (TGs), high-density lipoprotein cholesterol (HDL-C) and low-density

Peer Review Process: The Journal "Middle East Research Journal of Medical Sciences" abides by a double-blind peer review process such that the journal does not disclose the identity of the reviewer(s).

lipoprotein cholesterol (LDL-C). The study was approved by the Ethical Review Committee of Medical Faculty, Syiah Kuala University, Banda Aceh, Indonesia.

Gas Chromatography Analysis of Faecal SCFA Concentration

Stool samples were analysed for SCFA concentration with gas chromatography (GC) as described by a previous method (8). The amounts of acetate, propionate, butyrate, and valerate acids have been reported as mg/ml and %.

Statistical Analysis

Statistical analysis was carried out by SPSS, Results have been shown as mean ±standard deviation or median (interquartile range) for normally and nonnormally distributed continuous variables respectively, and number (percentages) for nominal variables. Shapiro-Wilk test was used to assess the normality of the variable's distributions. The Pearson correlation test (in the case of normal distribution) and Spearman's correlation test (in the case of non-normal distribution) were employed to examine the intensity and correlation between the two quantitative variables. All the tests were conducted at a significant level of 0.05.

RESULTS

A total of 50 patients joined in the study. Table 1 demonstrates the characteristics of the patients. Of these, 24 (48%) were male and the mean age of the studied population was 44.9 years old. The mean of fecal concentrations of acetate, propionate, butyrate, valerate, absolute butyrate and total SCFA were 58.13 %, propionate is 21.67 %, valerate is 2.34%, absolute butyrate is 2.34 mg/ml, and total SCFAs is 15.40 mg/dl, respectively.

Table 1: Characteristics of Subjects			
Variable	CSF (n=50)		
Age (years)	44.93 ± 11.13		
Sex, male female	24 (48%)/26 (52%)		
BMI (kg/m ²)	23,24±2,86		
SBP (mmHg)	140 (109 - 191)		
DBP (mmHg)	86 (63 - 148)		
Hemoglobin (g/dl)	14.30 (10.5 - 16.6)		
WBC (ul)	9900±2600		
Platelet (10 ³ /ul)	301 (150 - 406)		
Neutrophil-lymphocyte count ratio	1.78 (0.81 – 7.3)		
Total Cholesterol (mg/dl)	164 (117 – 297)		
LDL cholesterol (mg/dl)	107.27 ± 27.79		
HDL cholesterol (mg/dl)	47 (16 – 77)		
Triglyceride (mg/dl)	91 (72 – 269)		
Urea (mg/dl)	26.20 ± 10.4		
Creatinin (mg/dl)	1 (0.5 – 1.20)		
Random blood glucose (mg/dl)	93.60 ± 17.43		
Short Chain Fatty Acids (SCFA)			
Acetate Acids (%)	58.13 ± 7.29		
Propionate Acids (%)	21.67 ± 6.58		
Butyrate Acids (%)	12.2 ± 3.7		
Valerate Acids (%)	2.34 ± 0.98		
Absolute Butyrate Acids (mg/dl)	2.34 ± 1.26		
Total SCFAs (mg/dl)	15.40 ± 5.6		

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Data were presented as mean±SD, median (minimum-maximum) or n (%). BMI: Body Mass Index; SBP: Systolic Blood Pressure; DBP: Diastolic Blood Pressure; ESR: Erythrocyte Sedimentation Rate; LDL: Lipoprotein; HDL. High-density Low-density Lipoprotein. RCA: Right Coronary Artery; LCx: Left Circumflex Artery; LAD: Left Anterior Descending Artery; cTFC: Corrected Thrombolysis in Myocardial

Infarction Frame Count

Table 2 presents the association between shortchain fatty acids (SCFAs) with Lipid Profile. TC. LDL and TG not significantly correlated with SCFA but Pearson's correlation revealed a significant inverse correlation between HDL and butyrate acids (r = -0.532; p=<0.05) (figure 1).

Table 2: The associations between short-chain fatty acids (SCFAs) with Lipid Profile

		ТС	LDL	HDL	TG	
		r (P value)	r (P value)	r (P value)	r (P value)	
	Acetate Acids	0.063(0.823)	0.168(0.55)	0.325(0.237)	0.331(0.228)	
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Anil Batta., Middle East Res J. Med. Sci., Jan-Feb, 2024; 4(1): 31-35

0.081(0.773)	-0.327(0.234)	0.003(0.992)	-0.472(0.076)
-0,230 (0.410)	-0.188(0.503)	-0.532(0.041)*	-0.033(0.907)
-0.135(0.631)	-0.026(0.926)	-0.275(0.321)	0.409(0.130)
-0.280(0.312)	-0.437(0.103)	-0.1(0.722)	-0.394(0.146)
-0.108(0.701)	-0.354(0.196)	0.379(0.163)	-0.506(0.055)
	-0,230 (0.410) -0.135(0.631) -0.280(0.312)	-0,230 (0.410)-0.188(0.503)-0.135(0.631)-0.026(0.926)-0.280(0.312)-0.437(0.103)	-0,230 (0.410)-0.188(0.503)-0.532(0.041)*-0.135(0.631)-0.026(0.926)-0.275(0.321)-0.280(0.312)-0.437(0.103)-0.1(0.722)

*: p value correlation is significant at 0.05 level, r: Pearson's correlation coefficient.

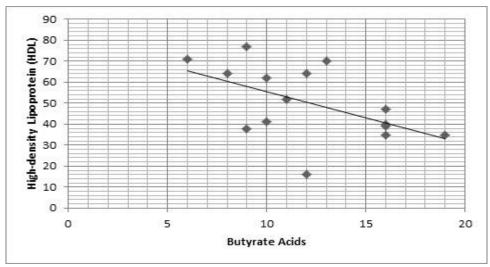


Figure 1: A negative correlation between butyrate acids with High-density Lipoprotein (HDL).

DISCUSSION

The gut microbiome is a growing topic of research in metabolic health and its link to CVD risk [9]. SCFAs are produced by the microbiota through the fermentation of ingestible polysaccharides and proteins, and are thought to represent the link between the microorganisms and the host. Individual SCFAs have also been shown to play a role in metabolism; for example, acetic acid supplementation reduces weight gain and improves glucose tolerance in obese people and diabetic rats [10], butyric acid protects against obesity and increases thermogenesis in mice [11], and propionic and butyric acids improve glucose homeostasis in mice [12]. By interacting with the diet, changes in the gut microbiota disrupt not only metabolism but also the composition of the host's lipids [13]. Dyslipidaemia has changed with SCFA, especially HDL. According to our findings, HDL is negatively correlated with butyrate acid in a patient with slow flow coroner.

Gut dysbiosis is generally characterized by a decrease in microbial population diversity and stability, and blooms in certain harmful bacteria [14]. Insulin resistance and abnormal levels of short-chain fatty acids (SCFAs) can occur from the metabolic network within the host harboring dysbiotic microorganisms being altered in situ [11]. The phylum Firmicutes produces butyric acid, the phylum Bacteroidetes produces propionic acid, and the majority of anaerobic bacteria make acetate [15].

Short-chain fatty acids appear to play a role in the regulation of fatty acid, glucose, and cholesterol

metabolism in cells. SCFAs can control lipolysis and adipogenesis. Endogenous lipolysis is inhibited by acetate and propionate, whilst extracellular lipolysis is regulated by propionate via a rise in lipoprotein lipase production, resulting in a decrease of the circulating lipid plasma levels and body weight [16, 17]. In rat colonic epithelial cells that convert SCFAs to acetyl-CoA, Zambell et al., discovered that acetate and butyrate are the predominant synthetic lipid substrates [18]. Finally, acetate, propionate, and butyrate appear to promote hepatic cholesterol uptake from the circulation, decreasing plasma cholesterol in model animal experiments. Furthermore, propionate inhibits cholesterol production effectively [19].

According to Granado-Serrano et al. participants with hypercholesterolemia had higher abundances of Odoribacter (Bacteroidetes) and Ruminococcus (Firmicutes) and lower abundances of Anaeroplasma (Tenericutes) and Haemophilus influenzae (Proteobacteria) [20]. Fu et al., found a negative connection between TG levels and the Pasteurellaceae genus [21]. Anaeroplasma abundance was also linked to an unfavorable lipid profile (IDL-C, TG-related biomarkers, and the ratio Total-C to HDL-C among others) [20]. The level of acetic acid in the feces was linked to IDL-C levels, which are linked to a more unfavorable lipid profile, but not propionic acid. Although there were no significant differences in serum levels of acetic and propionic acids between groups, hypercholesterolemia showed a profile with higher and lower abundance of acetic and propionic acids, respectively, than normocholesterolemia. These findings are in line with prior research that found that circulating

acetic acid stimulated "de novo" lipogenesis and cholesterogenesis in the liver, while propionic acid inhibited it [20-23]. There was no significant link between the lipid profile and the examined SCFA, acetate, propionate, and valerate in our study.

A study by Granado-Serrano et al., reported that there was no difference in the abundance of butyrate hypercholesterolemia feces between in and normocholesterolemia, and there was no association with any of the lipid biomarkers studied. However, in normocholesterolemia, its serum levels were greater, indicating a negative relationship with lipids associated with the worst profile, such as LDL-C. Total-C. LDL-TG, LDL-P (large and small), and Total-C to HDL-C ratio among others [20]. Butyrate has been shown to promote fatty acid production and cholesterogenesis in prior studies [19]. On the other hand, a study published by Gao Z et al., found that adding butyrate supplementation to the diet can help prevent other metabolic diseases including insulin resistance in rats, and this is linked to an energy expenditure and mitochondrial function activation pathway [11]. Butyrate levels were shown to be inversely linked with HDL in our study (p=0.05; r = -0.532).

CONCLUSION

In conclusion, the findings of this study show that dyslipidaemia, especially HDL level, is associated with butyrate acids in patients with slow-flow coroners. Further studies seem to be needed on other aspects of this relationship.

REFERENCES

- Beltrame, J. F., Limaye, S. B., & Horowitz, J. D. (2002). The coronary slow flow phenomenon–a new coronary microvascular disorder. *Cardiology*, 97(4), 197-202.
- Hawkins, B. M., Stavrakis, S., Rousan, T. A., Abu-Fadel, M., & Schechter, E. (2012). Coronary Slow Flow–Prevalence and Clinical Correlations– . *Circulation Journal*, 76(4), 936-942.
- 3. Chaudhry, M. A., Smith, M., Hanna, E. B., & Lazzara, R. (2012). Diverse spectrum of presentation of coronary slow flow phenomenon: a concise review of the literature. *Cardiology research and practice*, 2012.
- Marques, F. Z., Nelson, E., Chu, P. Y., Horlock, D., Fiedler, A., Ziemann, M., ... & Kaye, D. M. (2017). High-fiber diet and acetate supplementation change the gut microbiota and prevent the development of hypertension and heart failure in hypertensive mice. *Circulation*, 135(10), 964-977.
- Chen, W. J., Anderson, J. W., & Jennings, D. (1984). Propionate may mediate the hypocholesterolemic effects of certain soluble fibers in cholesterol-fed rats. *Proc Soc Exp Biol Med*, 175, 215–8.

- Kishimoto, Y., WAKABAYASHI, S., & TAKEDA, H. (1995). Effects of intravenous injection and intraperitoneal continual administration of sodium propionate on serum cholesterol levels in rats. *Journal of nutritional science and vitaminology*, 41(1), 73-81.
- Gibson, C. M., Cannon, C. P., Daley, W. L., Dodge Jr, J. T., Alexander, B., Marble, S. J., ... & Braunwald, E. (1996). TIMI frame count: a quantitative method of assessing coronary artery flow. *Circulation*, 93(5), 879-888.
- Chen, H. M., Yu, Y. N., Wang, J. L., Lin, Y. W., Kong, X., Yang, C. Q., ... & Fang, J. Y. (2013). Decreased dietary fiber intake and structural alteration of gut microbiota in patients with advanced colorectal adenoma. *The American of Clinical Nutrition*, 97(5), 1044-1052.
- Kelly, T. N., Bazzano, L. A., Ajami, N. J., He, H., Zhao, J., Petrosino, J. F., ... & He, J. (2016). Gut microbiome associates with lifetime cardiovascular disease risk profile among bogalusa heart study participants. *Circulation research*, 119(8), 956-964.
- Yamashita, H., Fujisawa, K., Ito, E., Idei, S., Kawaguchi, N., Kimoto, M., ... & Tsuji, H. (2007). Improvement of obesity and glucose tolerance by acetate in Type 2 diabetic Otsuka Long-Evans Tokushima Fatty (OLETF) rats. *Bioscience, biotechnology, and biochemistry*, 71(5), 1236-1243.
- Gao, Z., Yin, J., Zhang, J., Ward, R. E., Martin, R. J., Lefevre, M., ... & Ye, J. (2009). Butyrate improves insulin sensitivity and increases energy expenditure in mice. *Diabetes*, 58(7), 1509-1517.
- De Vadder, F., Kovatcheva-Datchary, P., Goncalves, D., Vinera, J., Zitoun, C., Duchampt, A., ... & Mithieux, G. (2014). Microbiota-generated metabolites promote metabolic benefits via gutbrain neural circuits. *Cell*, 156(1), 84-96.
- 13. Schoeler, M., & Caesar, R. (2019). Dietary lipids, gut microbiota and lipid metabolism. *Reviews in Endocrine and Metabolic Disorders*, 20, 461-472.
- Zeng, M. Y., Inohara, N., & Nuñez, G. (2017). Mechanisms of inflammation-driven bacterial dysbiosis in the gut. *Mucosal immunology*, 10(1), 18-26.
- Machiels, K., Joossens, M., Sabino, J., De Preter, V., Arijs, I., Eeckhaut, V. (2014). A decrease of the butyrate-producing species Roseburia hominis and Faecalibacteriumprausnitzii defines dysbiosis in patients with ulcerative colitis. *Gut*, 63, 1275–83.
- Lee, S. H., & Hossner, K. L. (2002). Coordinate regulation of ovine adipose tissue gene expression by propionate. *Journal of Animal Science*, 80(11), 2840-2849.
- Al-Lahham, S. A., Roelofsen, H., Rezaee, F., Weening, D., Hoek, A., Vonk, R., & Venema, K. (2012). Propionic acid affects immune status and metabolism in adipose tissue from overweight subjects. *European journal of clinical investigation*, 42(4), 357-364.

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- Zambell, K. L., Fitch, M. D., & Fleming, S. E. (2003). Acetate and butyrate are the major substrates for de novo lipogenesis in rat colonic epithelial cells. *The Journal of nutrition*, 133(11), 3509-3515.
- Demigné, C., Morand, C., Levrat, M. A., Besson, C., Moundras, C., & Rémésy, C. (1995). Effect of propionate on fatty acid and cholesterol synthesis and on acetate metabolism in isolated rat hepatocytes. *British journal of nutrition*, 74(2), 209-219.
- Granado-Serrano, A. B., Martín-Garí, M., Sánchez, V., Riart Solans, M., Berdún, R., Ludwig, I. A., ... & Serrano, J. C. E. (2019). Faecal bacterial and shortchain fatty acids signature in hypercholesterolemia. *Scientific Reports*, 9(1), 1772.
- Fu, J., Bonder, M. J., Cenit, M. C., Tigchelaar, E. F., Maatman, A., Dekens, J. A., ... & Zhernakova, A. (2015). The gut microbiome contributes to a substantial proportion of the variation in blood lipids. *Circulation research*, *117*(9), 817-824.
- Weitkunat, K., Schumann, S., Nickel, D., Kappo, K. A., Petzke, K. J., Kipp, A. P., ... & Klaus, S. (2016). Importance of propionate for the repression of hepatic lipogenesis and improvement of insulin sensitivity in high-fat diet-induced obesity. *Molecular nutrition & food research*, 60(12), 2611-2621.
- Wong, J. M., De Souza, R., Kendall, C. W., Emam, A., & Jenkins, D. J. (2006). Colonic health: fermentation and short chain fatty acids. *Journal of clinical gastroenterology*, 40(3), 235-243.